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REDUCTION OF BICYCLO[3.2.0]HEPT-2-EN-6-ONE AND 7,7-DIMETHYLBICYCLO[3.2.0]HEPT-2-EN-6-ONE USING DEHYDROGENASE ENZYMES AND THE FUNGUS MORTIERELLA RAMANNIANA

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SUMMARY: Bicyclo[3.2.0]hept-2-en-6-one (1) was reduced with an alcohol dehydrogenase from Thermoanaerobium brockii and a whole cell system (M. ramanniana) with excellent substrate enantioselectivity: 7,7-dimethylbicyclo[3.2.0]hept-2-en-6-one (2) was similarly reduced using the 3α,20β-hydroxysteroid dehydrogenase from Streptomyces hydrogenans while M. ramanniana furnished both 65-alcohols (4a), (6b) with high optical purity.

The synthetic utility of bicyclo[3.2.0] heptenones has been recognised for some time¹. In recent years this class of compound has been used as a starting point for the synthesis of a wide range of natural products including boonein², hirsutic acid-C³, hybridalactone⁴, multifidene⁵, pentalene⁶, pentalenolactones E and F⁷, prostaglandins^{8,9}, and the ophiobiolin skeleton¹⁰.

In order to enhance the usefulness of the bicyclo[3.2.0]-ring system for the preparation of <u>optically active</u> natural products, a general method of resolution of these bicyclic ketones would be valuable. Over the years, a number of procedures have been employed to prepare bicycloheptanones in optically active form¹¹. However, none of these methods can be used to prepare a <u>wide range</u> of chiral bicycloheptanones with good optical purity. We have investigated the use of dehydrogenase enzymes and a microorganism for stereoselective and enantioselective reduction of two bicycloheptenones.

Bicyclo[3.2.0]hept-2-en-6-one (1) was reduced to the corresponding <u>6endo-alcohol¹²</u> by three dehydrogenase enzymes (Table). Isolation of this alcohol from the reactions catalysed by horse liver alcohol dehydrogenase (HLAD) and 3α ,20 β -hydroxysteroid dehydrogenase (3α ,20 β -HSD) gave material with low optical rotation. In contrast, incubation of the substrate (1) with an alcohol dehydrogenase from <u>I. brockii</u> (Tab.) gave alcohol (3b) in high optical purity as assessed by optical rotation $[\alpha]_D^{21} = +69^0$ (c, 1.2 CHCl₃)¹³ and by spectroscopy after formation of Mosher's ester¹⁴. Unreacted ketone was also found to be optically active¹⁵.

50	7	8	

TABLE

Substrate	Dehydrogenase Enzyme	Cofactor (Recycling Agent)	Alcohol Produced	Predominant Enantiomer ^a 〈Enantioselectivity〉
1	HLAD (E.C. 1.1.1.1)	NADH (EtOH)	3	3b (<10% e.e.)
1	3α,20β-H5D (E.C. 1.1.1.53)	NADH (glucose/glucose dehydrogenase)	3	3b (<10% e.e.)
1	TabAD (E.C. 1.1.1.1)	NADPH (propan-2-ol)	3	3b (>95% e.e.)
2	HLAD	NADH (EtOH)	No reaction	_
2	TabAD	NADPH (propan-2-ol)	No reaction	-
2	3α,20β-HSD	NADH (HLAD, EtOH)	4	4a (>95% e.e.)
2	3α,20β-HSD	NADH (glucose/glucose dehydrogenase)	4	4a (>95% e.e.)

Reduction of Bicyclo[3.2.0]hept-2-en-6-one (1) and 7,7-Dimethylbicyclo[3.2.0]hept-2-en-6one (2) using Some Dehydrogenase Enzymes

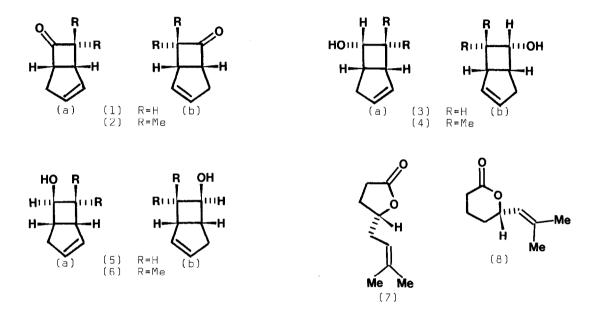
^aAfter 10-20% conversion

7,7-Dimethylbicyclo[3.2.0]hept-2-en-6-one (2) was unaffected by HLAD and the dehydrogenase from Tab. 3α ,206-HSD catalysed the reaction of (2) to a single product, identical by t.l.c. and h.p.l.c. to an authentic sample of the <u>6endo-alcohol</u> (4)¹⁶. A preparative scale reaction gave, after column chromatography, optically pure alcohol (4a) $[\alpha]_D^{21}$ -145°. The optical purity was assessed by g.l.c. of the isopropylurethane derivative over a chiral stationary phase¹⁷ and the absolute configuration was established by conversion into the lactone (7)¹⁸.

We have shown previously¹⁹ that the fungus <u>M. ramanniana</u> reduces one enantiomer of bicycloheptenone to give the <u>endo-alcohol</u> (3b) (e.e. = 90%) and recovered ketone (la) (e.e. = 80%). Incubation of <u>M. ramanniana</u> (ca. 125 g wet wt./litre) with the 7,7-dimethylbicycloheptenone (2) (1-2 g/litre) gave not only the corresponding <u>6endo-alcohol</u> (4) but also the <u>6exo-alcohol</u> (6). At low substrate concentrations (1-2 g/ litre) the rate of reduction of 7,7-dimethylbicycloheptenone was 50-70% the rate of reduction of the unsubstituted bicycloheptenone. At a higher substrate concentration (5 g/ litre), reduction of the ketone (2) proceeded at a markedly slower rate and only the <u>6endo-alcohol</u> was detected. A large scale run using freshly prepared <u>M. ramanniana</u> (ca. 200 g wet wt./litre) and dimethylbicycloheptenone (1 g/litre) gave a 50% total yield of equal quantities of the alcohols and g.l.c. analysis as described above showed that the endo-alcohol contained the

enantiomers (4a) and (4b) in the ratio 9:1. The <u>exo</u>-alcohol was of high optical purity $[\alpha]_D^{23} = \pm 109^0$ (c, 1.1 CHCl₃) with the enantiomer (6b) predominant (>95%). The absolute configuration of the alcohol (6b) was established by conversion into the lactone (8)¹⁸.

The lactones (7) and (8) have been used to prepare the optically active natural products eldanolide and leukotriene- B_4 respectively¹⁸, while the alcohol (3a) has been used to prepare prostaglandin- F_2^{α} in the naturally occurring configuration²⁰.



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- 16. Sodium borohydride reduction of ketone (2) gave two products in the ratio 10:1. The major, less polar product was assigned as the 6endo-alcohol (4) ν_{max} 3420 cm⁻¹; δ (CDCl₃) <u>inter alia</u> 3.82 (1H, d, J_{6 5} 8.2Hz, H-6), 2.6 (1H, m, H-5). The more polar product was assigned as the 6exo-alcohol (6) ν_{max} 3400 cm⁻¹; δ (CDCl₃) <u>inter alia</u> 3.5 (1H, d, J_{6 5} 3.5Hz, H-6), 2.6 (1H, m, H-5).
- 17. G.l.c. over a Chirasil-Valine column at 110⁰C with a helium flow of 2.0 ml/min.
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